

(1)

()

(NaCl)

.()

(mM NaCl 20)

(mM NaCl 200)

()

.(3 6)

.() ()

Saline signal

.(-0.3 Mpa PEG-6000) Osmotic signal

:

(1)

.2440 . . ()

Assessment the Importance of Induction and its Nature in Improving the Tolerance of Some ACSAD Durum and Bread Wheat Landraces to Salinity Stress

Ayman Shehada AL-Ouda⁽¹⁾

ABSTRACT

Several laboratory experiments were conducted to evaluate the response of ten ACSAD durum and bread wheat landraces to salinity stress tolerance at seedling stage. The salinity induction response technique was applied to assess the genetic variability, and the effect of induction nature in improving the tolerance to lethal levels of some abiotic stresses (drought and salinity). The applied screening tool was rapid and efficient in assessing the genetic variability for the response of studied durum and bread wheat landraces to salinity stress tolerance.

In general, induction enhanced the seedling tolerance to lethal salinity levels. The pre-exposure of seedlings to the optimum salinity stress level (20 mM NaCl) was associated with higher recovery growth after being exposed to the lethal level of stress (200 mM NaCl) compared with the non-induced seedlings. The recovery growth rate was as a consequence considered as an important biological parameter determining the degree of salinity stress tolerance. The step-wise transfer of seedlings to the lethal levels of stresses plays a pivotal role in assessing the potentiality of wheat salinity stress tolerance. Significant genetic variation occurred among durum and bread wheat landraces. But the tolerance of durum wheat landraces was at this phynological stage significantly higher than that of bread wheat. ACSAD durum and bread wheat landraces performed significantly better than the local cultivated varieties (Cham₆ and Cham₃). Significant differences were found in response to induction nature (salinity induction or osmotic induction), and stress type (different or identical). The salinity induction signal (20 mM NaCl) was more efficient in improving seedling tolerance than the osmotic induction signal (-0.3 Mpa PEG-6000) when the seedlings were transferred to the lethal levels of both salinity (200 mM NaCl) and drought (-1.5 MPa PEG-6000). The performance of both durum and bread wheat landraces was substantially better under the identical salinity stress compared with the different salinity stress and identical osmotic stress.

Generally, the salinity stress tolerance was found to be highly correlated with the nature of exposure (gradual or sudden), induction nature, and phynological stage and plant species.

Key words: Screening technique, Genetic variability, Induction nature, Salinity stress, Osmotic stress, Different stress, Identical stress, Landraces, ACSAD.

⁽¹⁾ Associate Professor, Agronomy Dept., Faculty of Agriculture, Damascus University, Damascus, Syria.
A cooperative expert, Plant Resources Division, Arab Center for the Studies of Arid Zones and Dry Lands (ACSAD). P.O. Box: 2440, Damascus, Syria.

20%

) (/ 2735) (4913) (1796)
. (2003)

.()

Soluble salts

.(Massoud, 1977)

954

)

.(2000)

...

%20

(1996 ,Alam ;1994 ,Sharma)

(Halophytes)

(Epestein et al., 1980)

(Francois, 1987)

(Prisco and O'leary, 1970)

Moisture stress

(Prisco *et al.*, 1981)

(Francois *et al.*, 1986)

.(Gupta and Srivastava, 1990)

.(Greenway and Munns, 1980)

.(Delane *et al.*, 1982)

Na⁺

Halophytes

Greenway (1973)

.(Pearson *et al.*, 1966) Mesophytes

Na⁺ Cl⁻

Water potential

()

LEA

)
(...

()
(Ganesh Kumar *et al.*, 1998; AL-Ouda, 1999)

Genetic variability

()

Sub-lethal level ()

Screening technique -1

-2

-3

()

Osmotic

-4

.stress

()

:

:

50%

%50

()

(NaCl

75 100 125 150 175 200 225 250 275)

NaCl

. 72

:(AL-Ouda, 1999)

$$100 \times \frac{\quad / \quad - \quad / \quad}{\quad / \quad} = (\%)$$

. 50%

:

16 (NaCl 0 20 30 40 50)

48

. 72

()

()

:

:

16

(-0.3 MPa)

48

()

72

48

72

() (-1.5 MPa)

:
16

16

(20 Mm NaCl)

()

72

48

16

() (200mM NaCl)

48

()

72

(NaCl)

(1)

200 mM NaCl

.48.04%

(NaCl)

Soluble salts

()
Water Potential gradient

.(Cossgrove, 1989)
.Osmotic Stress

.()

.(Cheeseman, 1988)

Specific Ion Toxicity
Cl⁻ Na⁺
(Cramer *et al.*, 1988)

Dry matter (Cheeseman, 1988)
(Sharma, 1996)

(1)

(%)	(Cm)	(%)	(Cm)	(mM NaCl)
-	62	-	42	
46.13	33.3	50.5	20.6	
5.88	58.2	4.73	40	20
14.05	53.2	14.07	36	30
21.12	48.8	23.31	32.2	40
29.74	44.8	28.97	29.6	50
5.087	3.257	3.857	1.674	L.S.D (5%)
14.35	3.59	10.48	2.75	C.V(%)

(2)

20 mM NaCl

4.73%

() 0.0 MPa

50.5%

(200 mM NaCl)

46.13%

(20 mM NaCl)

LEA

(Dasgupta and Bewley, 1984)

.Abiotic stresses

Amazallag *et al.*, (1990)

()

)

Amazallag *et al.*, 1990; AL-Ouda, (..

.(1999)

.(Mass and Grieve, 1992)

(2)

(%)	(Cm)	(%)	(Cm)	(mM NaCl)
15.12	49.4	18.33	33.4	75
15.97	48.9	16.32	34.3	100
15.97	48.9	17.07	33.75	125
26.80	24.6	34.63	26.8	150
29.03	41.3	35.28	26.5	175
42.18	33.3	48.04	21.3	200
48.28	30.1	54.63	18.6	225
55.32	26	63.96	14.8	250
63.40	21.3	72.43	11.3	275
5.087	3.257	3.857	1.674	L.S.D (5%)
14.35	3.59	10.48	2.75	C.V(%)

(3)

ACSAD1263

(13.8%) ACSAD 1239 (5.91%) (8.96%)
 (%12.80 %20.10) (10.44%)

(50.70% 58.80%) ACSAD1255 :
 (42.30% 50.90%) ACSAD1235 (%41.20 %51.80) ₃
 .(31.90% 40%) ACSAD1259
 .(3)

) ₃

(

Diffusion (CO₂) (Stroma) (CO₂/H₂O)

(3)

(%)	()	(%)	()	(ACSAD)
23.5	58.77	31.6	39	1231
42.3	43.3	50.9	29.85	1235
30.78	50.23	36.08	33.3	1237
10.44	45.33	13.8	31.11	1239
29.9	46.66	37.12	31.66	1245
50.7	49.55	58.8	35.44	1255
31.9	43.08	40	29.36	1259
22.2	56.42	35.5	38.42	1261
5.91	44.6	8.96	28.57	1263
23.6	40.42	31.6	25.14	1265
12.80	67.71	20.10	48.00	() 1105
41.20	41.50	51.80	25.50	() ₃
5.723	2.913	6.772	2.426	(0.05) L.S.D
13.86	3.46	12.87	4.23	C.V(%)

(4)

(25.70% 32.00%) ACSAD1063 (18.50% 23.70%) ACSAD1073

ACSAD1075 (52.90% 63.70%) ACSAD1069
 (42.14% 50.08%) ACSAD1065 (43.90% 54.80%)

(42.34% 48.50%) ACSAD1059

(%60.80) ()₆
 3) (%53.20) (4

(28.14% 34.73%)

37.02% 44.02%)

.(

(37.02%)

(28.14%)

(4)

(%)	()	(%)	()	(ACSAD)
35.5	44.4	42.5	30.3	1041
39.3	50.9	42.5	38.2	1047
42.34	43.3	48.5	30.7	1059
33.5	53.9	39.2	39.5	1061
25.7	52	32	36.3	1063
42.14	43.8	50.08	29.1	1065
52.9	37.66	36.7	23	1069
38.5	37.5	43.3	25.1	1071
18.5	45.75	23.7	32.5	1073
43.9	35.4	54.8	22.3	1075
53.20	36.50	60.80	24	₆
7.305	3.365	7.622	2.517	L.S.D (0.05)
1.49	4.41	10.09	4.75	C.V(%)

(5)

-

-

(-)

(-)

ACSAD 1235

ACSAD 1261

ACSAD 1231

ACSAD 1239

(-)

(-)

(-)

(-)

ACSAD 1259

(14.65%) ACSAD 1245

ACSAD 1261 (%19.97) ACSAD 1231 (%21.18)

ACSAD 1235

(42.25%) ACSAD 1239 (%36.55) ACSAD 1237

(%32.10)

(-)

-)

(-)

(

ACSAD

(56.42)

1261

ACSAD 1259

(68.52) ACSAD 1231

(60.70)

ACSAD (%92.60) ACSAD 1237

(%99.30) ACSAD 1239

(%79.30) 1235

(5)

(%)	(%)	(%)	()	(%)	()		
81	65	21.6 17 12 10	63.5 67.6 72 71	25.6 23 13 18.3	4803 49.8 53.3 53.2	.	ACSAD 1231
79.3	62.6	41 28.6 30.3 21	52.2 56.6 53.3 62.5	39.2 35 30.6 23.6	38 41.6 43.3 47.6	.	ACSAD 1235
92.6	75.6	28.8 23.6 39.3 29.6	65.8 70.8 55.8 65	35.5 28.6 47.6 34.5	48.8 53.8 39.6 49.3	.	ACSAD 1237
99.3	83.3	44 34.3 32.6 33	55.6 65 66.6	49 42.5 39.3 38.2	42.5 47.6 50.3 51.3	.	ACSAD 1239
59.3	44.6	19.6 23.8 2.6 5.3	47.6 45.2 59.2 57	22.3 29 4 3.3	34.6 31.6 44.4 44.3	.	ACSAD 1245
57.3	43	17.2 25 8 23	47.2 41.5 69.2 58	24.2 38 7.8 23.8	32.5 26.6 53.2 43.8	.	ACSAD 1255
75.2	57.6	20.2 26 8 23	60.03 55.6 69.2 58	23.3 32.3 7.8 23.8	44.2 39 53.2 43.8	.	ACSAD 1259
58.6	45	6.8 6.6 15.4 23.6	54.6 56.6 57 44.5	12.6 10.3 14.5 30.3	39.3 40.3 41.5 31.3	.	ACSAD 1261
77	61	14.6 42 29 35.3	65.6 44.3 54.6 49.6	20.6 48.6 36.8 41.2	48.6 29.6 38.6 36.2	.	ACSAD 1263

(-)

(-)
 (-)
 (-)
 (-)

(5)

C.V(%)	(L.S.D)			
	0.01	0.05		
7.24	2.221	1.673		
	3.332	2.509		
	6.663	5.019		
20.74	4.033	3.038		
	6.050	4.557		
	12.10	9.114		
5.38	1.692	2.246		
	2.538	3.369		
	5.076	6.739		
20.73	3.403	2.563		
	5.105	3.845		
	10.21	7.690		

(20mM Na Cl)

(-0.3MPa)

Induced-salt

.Induced-osmotic signal

signal

(6)

(-)

(-)

38)

ACSAD 1059

(-)

(51.3 67)

(54

(-)

(45.35)

(32.17)

ACSAD 1071

(6) (36.62)

(24.20)

(6)

(-)

(-)

(-)

(-)

(-)

(-)

(6)

(%)	(%)	(%)	()	(%)	()		
62	46.3	45.6 62 6.7 43.5	33.6 23.5 56.8 35	50 70.6 8.2 51.6	21 13.3 43.3 22.3	.	ACSAD 1041
65	55.6	54.6 55.2 24.8 42.6	39.3 32 53.6 40.8	49.3 61.5 20.3 47.8	28 21.3 44 29	.	ACSAD 1047
72.3	58.6	58 54.6 14.5 29	30.3 32.8 67 51.3	70.6 67.3 16.6 34.5	17.2 19.2 54 38.3	.	ACSAD 1059
66	49.6	41.3 54.2 10.6 38.2	38.6 30.2 57.8 40.5	47 63.5 11.6 36.2	26.3 18 43.6 31.6	.	ACSAD 1061
69.6	54.3	56.6 49.3 16 18.8	30.2 35 58.3 56.3	69 62.6 22.3 23.8	16.6 20 42 41.2	.	ACSAD 1063
76	61.3	46.3 59.3 27.3 50	40.6 30.8 55.2 38	55 68.3 32.3 59.6	27.5 19 41.3 24.6	.	ACSAD 1065
37.3	30.6	22.3 27.8 25.6- 4.1-	28.8 26.8 46.6 38.6	29 32.3 9.4- 12.2	17.5 16.6 33.5 26.3	.	ACSAD 1069
57.8	43.3	33.2 38 40.3 35	33.2 38 40.3 35	52 39.6 35.5 48.8	20.8 25.8 28 22.2	.	ACSAD 1071
62	48.6	35 30 68.5 64.8	35 30 68.5 64.8	54 58 12.1- 0.98	21.3 19.3 52 51.2	.	ACSAD 1073
47.6	35.6	23.8 23.6 51.3 45.2	23.8 23.6 51.3 45.2	59.6 60.5 8.97- 7.2	14.3 14 38.8 33	.	ACSAD 1075

(6)

C.V(%)	(L.S.D)			
	0.01	0.05		
7.99	1.550	1.169		
	2.451	1.848		
	4.903	3.697		
13.30	3.544	2.673		
	5.604	4.226		
	11.21	8.457		
7.29	2.038	1.537		
	3.222	2.430		
	6.445	4.859		
18.26	4.029	3.038		
	6.370	4.803		
	12.74	9.606		

1

Genetic Variability

2

-3

/ () 4

()
5

REFERENCES

.2003

.2000

- Alam, S.M. (1994). Nutrients uptake by plants under stress conditions (In Handbook of Plant and Crop Stress), pp. 233-236, Marcel Dekker, Inc. New York.
- AL-Ouda, A. (1999). Genetic variability in temperature tolerance in sunflower hybrids: An assessment based on physiological and biochemical traits. Ph. D. Thesis Submitted to Crop Physiology Dep., UAS, India.
- Amzallag, G.N.; Lerner, H.R. and Poljakoff-Mayber, A. (1990). Induction of increased salt tolerance in sorghum bicolor by NaCl pretreatment. J. Exp. Bot., 41: 29-34.
- Cheeseman, J. M. (1988). Mechanism of salinity tolerance in plants. Plant Physiol., 87: 547-550.
- Cossgrave, D. J. (1989). Characterization of long term extension of isolated cell walls from growing cucumber hypocotyls. Planta, 177: 121.
- Cramer, G. R; Epstein, E. and Lauchli, A. (1988). Kinetics of root elongation of maize in response to short-term exposure to NaCl and elevated calcium concentration. J. Exp. Bot., 39: 1513.
- Dasgupta, J. and Bewley, D. (1984). Variation in protein synthesis in different regions of greening leaves of barley seedlings and effects of imposed water stress. J. Exp. Biol., 35: 1450.
- Delanne, R.; Greenway, H.; Munns, R. and Gibbs, J. (1982). Ion concentration and carbohydrate status of the elongating leaf tissue of *Hordeum vulgare* growing at high external NaCl. J. Exp., Bot., 33: 557-573.
- Epstein, E.; Norlyn, J. D.; Rush, D.W.; Kingsbury, R.W.; Kelley, D.W.; Gunningham, G. A. and Wrona, A. F. (1980). Saline culture of crops: A genetic approach, Science, 210: 399-404.
- Epstein, E. (1985). Salt tolerance crops: origin, development and prospects of the concept, Plant Soil., 89: 187-198.
- Frnacois, L.E. (1987). Salinity effects on asparagus yield and vegetative growth, J.Am. Soc. Hort. Sci., 112:432-436.
- Francois, L. E.; Mass, E.V.; Donovan, T. J. and Youngs, V. L. (1986). Effect of salinity on grain yield and quality, vegetative growth and germination of semi-dwarf and durum wheat, Aron. J., 78: 1053-1058.

- Ganesh Kumar; Krishna Prasad, B.T.; Savita, M.; Gopala Krishna, R.; Mukhopdyay, K.; Rama Mohan, G. and Udaya Kumar, M. (1998). Enhanced expression of heat shock proteins in thermo-tolerant lines of sunflower and their progenies selected on the basis of temperature induction response. *Theor. Appl. Genet.* On 28th October, 1998.
- Greenway, H. and Munns, R. (1980). Mechanisms of salt tolerance in non-halophytes, *Annu. Rev. Plant Physiol.*, 31: 149-190.
- Greenway, H. (1973). Salinity, plant growth and metabolism, *J. Aust. Inst. Agr. Sci.*, 39: 24.
- Gupta, S.C. and Srivastava, J.P. (1990). Effect of salt on morpho-physiological parameters in wheat (*Triticum aestivum L.*) *Indian J. Plant Physiol.*, 32: 169.
- Mass, E.V. and Grieve, C.M. (1992). Salt tolerance of plants at different stages of growth, *Proc. Int. Conf. on Current Development of Salinity and Drought Tolerance of Plants*, January 7-11, 1990, Tandojam, Pakistan.
- Massoud, F.I. (1977). Basic principles for prognosis and monitoring of salinity and sodicity. *Proc. Int. Salinity Conf. Texas technical University, Lubbock*, pp. 432-454.
- Pearson, G.A.; Ayers, A.D. and Eberhard, D.L. (1966). Relative salt tolerance of rice during germination and early seedling development. *Soil Sci.*, 102: 151-156.
- Prisco, T.J. and O'Leary, J.W. (1970). Response of osmotically-stressed plants to growth regulators. *Adv. Front Plant Sci.*, 25: 129.
- Prisco, J.T. and O'Leary, J.W. (1970). Osmotic and toxic effects of salinity on germination of *Phaseolus vulgaris L.* seeds. *Turriabxla*, 20: 174-184.
- Prisco, J.T.; Filho, J.E. and Filho, G.E. (1981). Effect of NaCl salinity on cotyledon starch mobilization during germination of *Vigna unguiculata Walp.* *Seed, Review Brazil, Bot.*, 4: 63-71.
- Sharma, S.K. (1996). Soil salinity effects on transpiration and net photosynthetic rates, stomatal conductance and Na⁺ and Cl⁻ contents in durum wheat. *Biologia Plantarum*, Vol. 38: 519-523.

Received	2005/12/15	
Accepted for Publ.	2006/03/12	